PACKING INSTRUCTIONS–WET PACK
31XX CAPILLARY ARRAY, 2-16 CAPS

When sending capillaries to be regenerated, include all original packaging materials. This would include adding distilled water to the bottle and in WetPack (see below).

PREPARING YOUR ARRAY FOR REGENERATION:

1. FOR ARRAYS WHICH WILL BE PULLED FROM THE INSTRUMENT AND SHIPPED DIRECTLY:

A. Remove array from ABI genetic analyzer.

B. Fill glass bottle with ddH2O and assemble capillary end with o-ring and screw cap (figure 1). Note: make sure screw cap is tight. For 3730 arrays, make sure the bottle is attached firmly to frame; otherwise the bottle can swing freely and damage capillaries.

C. Place array in its original shipping container (figure 2) and note the serial number, date removed, and number of runs on the array. In many cases the box will have a different serial number than the array, so make sure you read the number from the array itself.

D. After array has been correctly packaged, ship the array to Gel Company. If you pull an array on a Friday, store both ends of the array in ddH20 until Monday, then follow the instructions from step 1 above.

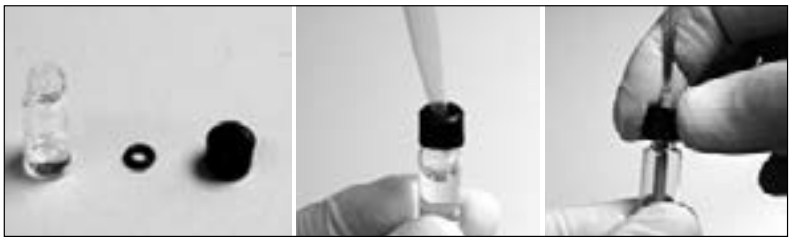

FIG. 1
Assemble bottle, o-ring and bottle cap together. With the cap loose, fill bottle with distilled water. Insert wet capillary end into bottle assembly without cutting o-ring. This is done by wobbling the capillary rod as it is going thru o-ring. Once inside, tighten the cap so that the o-ring is squeezed enough to seal water filled bottle.

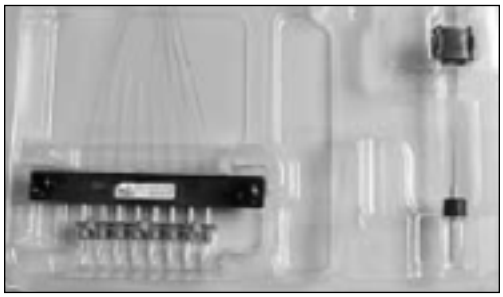

FIG. 2
50cm or longer to be shipped in 180º position (as shown). 36cm to be shipped in 90º position (not shown). Please include original capillary cover taped securely to container for return shipment.

2. FOR ARRAYS WHICH HAVE BEEN REMOVED PREVIOUSLY AND ARE IN STORAGE:

A. Whenever you store an array off the machine, and the array contains polymer, make sure both ends of the array are stored wet (both ends submersed in ddH20).

B. One convenient way to store the capillary ends wet is the use of a WetPack. A WetPack is simply a 16 well cut out of a flat 96 well tray (catalog# MAP3196C) and a 16 cap cut out of a septamat (Catalog #SEP96-20). Fill the 16 well cut tray with ddH20, attach the cut septamat, then secure the two pieces together with three rubber bands (please see figure 3). Next, insert the capillary ends through the septamat. This method is nice in that you don't waste a lot of water and water does not splash over the array during shipment. Also, the WetPack will fit when placing the array in its original packaging. This step is not required, but it does generally produce the best results for array regeneration.

C. Follow steps 1A to 1D above.

FIG. 3
Fill the tray wells with distilled water, insert rubber septa mat, wrap tightly with rubber bands. Insert capillary array into the assembled Wet Pack.

PACKING INSTRUCTIONS–WET PACK
3730/3730 XL CAPILLARY ARRAY, 48-96 CAPS

When sending capillaries to be regenerated, include all original packaging materials. This would include adding distilled water to the bottle and in WetPack (see below).

PREPARING YOUR ARRAY FOR REGENERATION:

1. FOR ARRAYS WHICH WILL BE PULLED FROM THE INSTRUMENT AND SHIPPED DIRECTLY:

A. Remove array from ABI genetic analyzer.

B. Fill glass bottle with ddH2O and assemble capillary end with o-ring and screw cap (figure 1). Note: make sure screw cap is tight. For 3730 arrays, make sure the bottle is attached firmly to frame; otherwise the bottle can swing freely and damage capillaries.

C. Place array in its original shipping container (figure 2) and note the serial number, date removed, and number of runs on the array. In many cases the box will have a different serial number than the array, so make sure you read the number from the array itself.

D. After array has been correctly packaged, ship the array to Gel Company. If you pull an array on a Friday, store both ends of the array in ddH20 until Monday, then follow the instructions from step 1 above.

2. FOR ARRAYS WHICH HAVE BEEN REMOVED PREVIOUSLY AND ARE IN STORAGE:

A. Whenever you store an array off the machine, and the array contains polymer, make sure both ends of the array are stored wet (both ends submersed in ddH20).

B. One convenient way to store the capillary ends wet is the use of a WetPack. A WetPack is simply a 96 well tray (catalog# MAP3196C) and a 96 cap septamat (Catalog #SEP96-20). Fill the 96 well tray with ddH20, attach the septamat, then secure the two pieces together with three rubber bands (please see figure 3). Next, insert the capillary ends through the septamat. This method is nice in that you don't waste a lot of water and water does not splash over the array during shipment. Also, the WetPack will fit when placing the array in its original packaging. This step is not required, but it does generally produce the best results for array regeneration.

C. Follow steps 1A to 1D above.

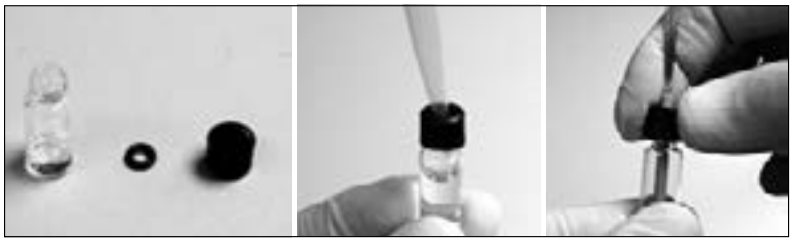

FIG. 1
Assemble bottle, o-ring and bottle cap together. With the cap loose, fill bottle with distilled water. Insert wet capillary end into bottle assembly without cutting o-ring. This is done by wobbling the capillary rod as it is going thru o-ring. Once inside, tighten the cap so that the o-ring is squeezed enough to seal water filled bottle.

FIG. 2
Please include original capillary cover taped securely to container for return shipment.

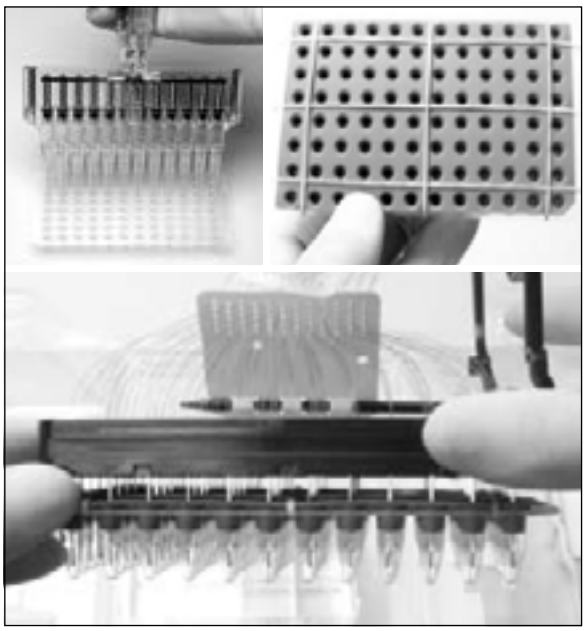

FIG. 3
Fill the tray wells with distilled water, insert rubber septa mat, wrap tightly with rubber bands. Insert capillary array into the assembled Wet Pack.